



Human Fertility

an international, multidisciplinary journal dedicated to furthering research and promoting good practice

ISSN: 1464-7273 (Print) 1742-8149 (Online) Journal homepage: <http://www.tandfonline.com/loi/ihuf20>

A blessing and a curse: is high NK cell activity good for health and bad for reproduction?

Sophie Templer & Gavin Sacks

To cite this article: Sophie Templer & Gavin Sacks (2016): A blessing and a curse: is high NK cell activity good for health and bad for reproduction?, Human Fertility, DOI: [10.1080/14647273.2016.1219072](https://doi.org/10.1080/14647273.2016.1219072)

To link to this article: <http://dx.doi.org/10.1080/14647273.2016.1219072>



Published online: 12 Aug 2016.



Submit your article to this journal [↗](#)



View related articles [↗](#)



View Crossmark data [↗](#)

REVIEW ARTICLE

A blessing and a curse: is high NK cell activity good for health and bad for reproduction?

Sophie Templer^a and Gavin Sacks^{a,b,c,d}

^aSchool of Women's and Children's Health, University of New South Wales, Sydney, Australia; ^bIVF Australia, Sydney, Australia; ^cSt George Hospital, Sydney, Australia; ^dRoyal Hospital for Women, Sydney, Australia

ABSTRACT

Few topics in recent reproductive medicine have been the subject of as much controversy, media attention and passionate debate as natural killer (NK) cells and their role in reproductive failure. The question of whether elevated NK cell levels are a cause of infertility and pregnancy loss, and whether they provide a potential target for therapy to improve reproductive outcomes, lacks a definitive answer. It is clear, however, that a significant number of women with reproductive failure have abnormal NK cell parameters reflecting high immunological activity. Amongst all the debate, the wider implications of NK cell overactivity – and attempts to suppress it – have not yet been considered. The literature suggests that although elevated NK cell activity may not be conducive to reproduction, it could in fact be beneficial in other areas of health and disease such as cancer and infection. Further research is needed to determine whether this hypothesis holds true in women with NK cell-related reproductive failure.

ARTICLE HISTORY

Received 26 June 2015
Accepted 3 January 2016

KEYWORDS

In vitro fertilisation; infertility; natural killer cells; reproductive failure; reproductive immunology

Introduction

Reproductive failure, a broad set of conditions encompassing infertility, recurrent miscarriage and recurrent implantation failure (RIF), is common but often poorly understood. Infertility, defined as the failure to conceive after at least 12 months of regular unprotected sexual intercourse, affects 14% of couples, while recurrent miscarriage (RM) – three or more consecutive pregnancy losses before 20 weeks' gestation – occurs in 1–3% of women (Gelbaya, Potdar, Jevé, & Nardo, 2014; King, Smith, Chapman, & Sacks, 2010). RIF has various definitions but is generally accepted as the failure of three or more *in vitro* fertilization (IVF) cycles where one or two high quality embryos are transferred in each cycle (Simon & Laufer, 2012). In a large proportion of patients within these groups – including 50% of women with RM and 15–30% of infertile couples – no explanation is found for their reproductive failure from traditional fertility investigations (Gelbaya et al., 2014; Quenby & Farquharson, 2006).

For these patients, many of whom have experienced the physical, emotional and financial stress of multiple failed cycles of IVF, the diagnosis (or lack thereof) of unexplained reproductive failure is often greatly unsatisfying. Many such patients are in a position to benefit (or at least believe they are) from NK

cell testing, an area of reproductive medicine with a long history but one that remains controversial. The extensive debate and politics surrounding NK cells and their relevance in reproductive failure has maligned the topic away from the mainstream (Sacks, 2015). Indeed, a recent survey of IVF clinicians revealed that 69% would recommend immunological investigations in patients with RM, although only 8% would consider a NK cell assay; similarly, 56% would recommend immunological testing in cases of RIF but only 9% would suggest a NK cell assay (Kwak-Kim et al., 2013).

Twenty years of research have provided a multitude of evidence to support the notion that NK cell overactivity is correlated with reproductive failure. One estimate suggests that elevated NK cell numbers or activity are found in 15–25% of women with unexplained RM (Sacks, 2014), but it is not yet clear what these findings mean or whether they can, so to say, explain the unexplained. No disease has yet been associated with elevated NK cell activity, but it has been suggested that such a disease does exist – one that is manifested in the failure of implantation and progression of pregnancy (Sacks, 2015). It is likely, if such a disease exists, that other manifestations reflective of high NK cell activity will be present. To look for these, it is first essential to determine the functions of

NK cells in other areas of health and disease, including autoimmunity, infection and cancer.

NK cells and fertility

It has long been assumed that the maternal immune system must be suppressed or redirected in some way to ensure a successful implantation and pregnancy. Investigations into the role of NK cells in infertility and miscarriage began in the mid-1990s, with the claim by Aoki et al. (1995) that RM patients with elevated peripheral NK (pNK) cell activity in the preconceptional period had an increased risk of miscarriage. Soon after, Roussev, Kaider, Price, and Coulam (1996) demonstrated that women with RM, RIF and infertility were more likely to have elevated pNK cell levels than fertile controls, while Beer, Kwak, and Ruiz (1996) claimed to be the first researchers to have shown that elevated pNK cell levels could predict reproductive failure in women with RM and RIF.

Many researchers over the past twenty years have sought to prove (or disprove) this hypothesis posed by early investigators. Significant correlations have been drawn between RM and various pNK cell parameters, including cell levels (both as a percentage of lymphocytes and as an absolute number), CD56dim levels, cytotoxicity, expression of the activation markers CD69 and CD16, and secretion of TNF- α and IFN- γ , all of which are increased in RM (Emmer et al., 2000; Fukui et al., 2008; Ghafourian, Karami, Khodadadi, & Nikbakht, 2014; Hadinedoushan, Mirahmadian, & Aflatounian, 2007; King et al., 2010; Lu et al., 2011; Ntrivalas et al., 2001; Ramos-Medina et al., 2013; Yoo et al., 2012). By contrast, RM is also associated with a decrease in certain pNK cell parameters, including CD56bright levels, CD56bright:CD56dim ratio, expression of the inhibitory marker CD94, and secretion of IL-4 and IL-10 (Emmer et al., 2000; Fukui et al., 2008; King et al., 2010; Ntrivalas et al., 2001). The majority of these observations are also true in RIF (Fukui et al., 2008; Ghafourian et al., 2014; Lu et al., 2011; Ramos-Medina et al., 2013; Sacks et al., 2012).

Decidual NK (dNK) and uterine NK (uNK) cells are more difficult to measure, but similar observations have been made: women with RM have higher uNK cell levels (Clifford, Flanagan, & Regan, 1999; Quenby, Kalumbi, Bates, Farquharson, & Vince, 2005; Tuckerman, Laird, Prakash, & Li, 2007), higher dNK cell levels (Almasry, Elmansy, Elfayomy, & Algaidi, 2015), and a lower CD56bright:CD56dim dNK ratio (Yamamoto, Takahashi, Kase, & Mori, 1999). Women with RIF have also been shown to have higher uNK

cell levels than controls (Fukui et al., 2008; Santillan et al., 2015).

It is not yet understood how NK cells contribute to reproductive failure, but a number of possible mechanisms have been proposed. Firstly, it has been suggested that NK cell overactivation results in excessive production of Th1-type cytokines, primarily TNF- α and IFN- γ (Fukui et al., 2008, 2011). The maternal immune response shifts from Th1 to Th2 dominance in normal pregnancy, but Th1 activity is enhanced in reproductive failure, suggesting that it creates a hostile or unhelpful environment for implantation and pregnancy (Fukui et al., 2011; Laird et al., 2003). Furthermore, pNK production of cytokines that are known to be conducive to pregnancy, including IL-4 and IL-10, is reduced in reproductive failure (Fukui et al., 2008). Secondly, impaired epitope matching between maternal inhibitory killer-cell immunoglobulin-like receptors (KIRs) and trophoblastic HLA-C, which reduces the inhibition of uNK-mediated trophoblastic cytolysis, is thought to contribute to overactivation and excessive cytolytic activity of uNK cells (Faridi & Agrawal, 2011; Varla-Leftherioti et al., 2005). Thirdly, since uNK cells contribute to endometrial angiogenesis and spiral artery remodelling, it has been suggested that increased uNK cell density causes increased peri-implantation blood flow and the early establishment of maternal circulation, resulting in excessive oxidative stress with the potential to trigger miscarriage (Quenby et al., 2009). Finally, high uNK cell levels are associated with decreased cortisol synthesis by decidualizing cells and impaired induction of local mineralocorticoid-dependent enzymes. These findings suggest that elevated uNK cell levels may be a useful marker for identifying women at risk of pregnancy loss due to uterine cortisol deficiency (Kuroda et al., 2013). As observed in the liver, NK cells also play an important role in facilitating tissue regeneration and reducing fibrosis (Krizhanovsky et al., 2008); similar processes in the endometrium could lead to reproductive failure as a result of aberrant decidualization and regeneration of endometrial tissue.

None of the proposed mechanisms for the role of NK cells in reproductive failure have been clearly substantiated, and it is also possible that NK cell overactivity is a marker of – rather than a cause for – reproductive failure. Yet it is clear from the evidence outlined above that a link does exist between NK cell activity and poor reproductive outcomes. Certain pNK cell parameters, particularly percentage and cytotoxicity, have been shown to be highly specific for RM (Coulam, Goodman, Roussev, Thomason, & Beaman, 1995; King et al., 2010; Lee et al., 2013) as well as

predictive of future pregnancy outcomes (Ramos-Medina et al., 2013). More importantly, though, NK cell testing facilitates the identification of patients who are likely to benefit from immunotherapy, and preliminary trials have demonstrated an improvement in rates of pregnancy and live birth in reproductive failure patients with elevated NK cell levels who are treated with intravenous immunoglobulin (IVIg), intralipid and/or prednisolone (Clark, 2008; Coulam & Acacio, 2012; Morikawa et al., 2001; Polanski et al., 2014). An additional, perhaps undervalued, benefit of NK cell testing is the reassurance it provides to patients that their doctor is seeking an explanation for (and solution to) their problem, as well as thinking beyond the unsatisfying diagnosis of ‘unexplained’ reproductive failure and, importantly, offering them hope (Sacks, 2014).

The practice of NK testing in the fertility setting remains controversial, however, as several groups have found no association between NK cell abnormalities and reproductive failure (Katano et al., 2013; Ozcimen, Kiyici, Uckuyu, & Yanik, 2009; Thum et al., 2005). More importantly, others have reported that no significant correlation exists between pNK and uNK cell levels (Junovich et al., 2013; Lukassen et al., 2004; Yamamoto et al., 1999), which has led some to conclude that any attempts to extrapolate information about the immune environment of the uterus from pNK cell tests are inherently flawed (Clark, 2008; Laird, Mariee, Wei, & Li, 2011).

The most likely reason for the markedly contradictory conclusions reached by different investigators is the vastly different methodologies used to measure NK cells. Not only are NK cells studied in different compartments of the body (the blood, endometrium or decidua), but the techniques used also vary between labs and even individual technicians (Laird et al., 2011; Sacks, 2015). Measuring uNK cells from an endometrial biopsy is particularly problematic since it is technically difficult and subjective (Sacks, 2015). The timing of sample collection (stage of menstrual cycle, during pregnancy or after miscarriage) is also crucial (Laird et al., 2003). Furthermore, a number of different parameters have been analysed, including percentages or absolute numbers of NK cells, cell subtypes and surface markers, with many different (often arbitrary) thresholds used to define normal and abnormal results (Laird et al., 2011). Numerous miscarriage studies have also failed to perform foetal karyotyping in every case, since chromosomally normal and abnormal miscarriages should be examined separately (Laird et al., 2003). With this many variables to account for differences in results, it is no wonder that researchers have arrived at such different conclusions.

NK cells and autoimmunity

NK cells have been implicated in multiple autoimmune diseases in both protective and pathogenic roles. The most consistent finding amongst patients with autoimmune disease is a low pNK cell count, which has been described in multiple sclerosis (MS), rheumatoid arthritis (RA), systemic lupus erythematosus (SLE), type I diabetes mellitus (T1DM), Sjögren’s disease, psoriasis and autoimmune thyroid disease. Rather than a true NK cell deficiency, however, this most likely reflects the sequestration of NK cells to target tissues, since NK cells accumulate in the affected tissues of RA (synovium), psoriasis (skin lesions), T1DM (pancreatic islets), alopecia areata (hair follicles), and dermatomyositis (muscle). Certain activating KIR alleles have also been identified as predisposing factors to a number of autoimmune diseases, while a lack of certain inhibitory KIRs predisposes to others. It is unclear whether changes in NK cell location and function in autoimmunity are a primary pathogenic defect, or whether they simply reflect a secondary response to the disease. Certain patterns in NK cell abnormalities, however – that they are often present in newly onset but not chronic disease and active but not quiescent disease – suggest that NK cells contribute to the initiation of autoimmune processes rather than being the result of them (Fogel, Yokoyama, & French, 2013).

Yet NK cells also appear to play a protective role in autoimmune disease. The production of IL-10 by NK cells inhibits dendritic cells (DCs), reducing antigen presentation and therefore T cell proliferation which might otherwise result in an autoimmune response. NK cells are also capable of lysing activated macrophages, thereby downregulating inappropriate phagocytic activity (Folci, 2014). The most convincing evidence for the protective role of NK cells in a specific disease is MS, since pNK cell numbers are diminished in active disease but are restored in remission and upon effective treatment with the monoclonal antibody dacluzimab (Flodstrom-Tullberg, Bryceson, Shi, Hoglund, & Ljunggren, 2009; Folci, 2014). Nevertheless, much like the area of reproductive failure, the evidence for NK cell involvement in autoimmunity is, as described by Fogel et al. (2013), ‘tantalising but incomplete’.

NK cells and cancer

NK cells play an important role in cancer immunosurveillance, primarily through their ability to kill tumour cells directly without prior sensitisation. This occurs via four mechanisms: (1) perforin- or granzyme-mediated

cytotoxicity; (2) death receptor-mediated apoptosis; (3) release of effector molecules such as IFN- γ ; and (4) antibody-dependent cell-mediated cytotoxicity (Cheng, Chen, Xiao, Sun, & Tian, 2013). Furthermore, NK cells and their products interact with other components of the immune system to initiate and maintain a powerful system-wide anti-tumour response. By communicating with DCs, NK cells improve antigen presentation and thereby facilitate the generation of a tumour-specific cytotoxic T cell response. This effect is compounded by the ability of NK cells to promote CD4⁺ and CD8⁺ T cell differentiation as well as induce T cell-mediated cytotoxicity through IFN- γ (Cheng et al., 2013; Langers, Renoux, Thiry, Delvenne, & Jacobs, 2012). Chemokines produced by NK cells also induce the migration of DCs and T cells to malignant tissue (Terunuma, Deng, Dewan, Fujimoto, & Yamamoto, 2008).

A landmark study by Imai, Matsuyama, Miyake, Suga, and Nakachi (2000) measured the baseline natural cytotoxicity levels of 3500 participants using a chromium-51 release assay of the peripheral blood, dividing the participants by tertiles into groups of low, medium and high cytotoxicity levels. The participants were followed up after an 11-year period to assess cancer incidence. Participants with low levels of cytotoxic activity had a significantly increased risk of cancer, while those in the medium and high cytotoxicity groups had a relative risk of 0.63 compared to the low-level group. This trend was especially marked in the female population; women with high cytotoxic activity had approximately half the risk of cancer as compared to women with low cytotoxicity.

The function of NK cells in cancer has been thoroughly reviewed by Levy, Roberti, and Mordoh (2011) and Cheng et al. (2013). The presence of NK cells in malignant tissue is a positive prognostic marker: high levels of tumour-infiltrating NK cells are associated with favourable outcomes in various carcinomas, while lower intraneoplastic NK cell counts are associated with a higher risk of metastasis (Cheng et al., 2013; Levy et al., 2011). Cancer patients also appear to have an altered NK cell phenotype, marked by overexpression of inhibitory receptors, poor expression of activating receptors, increased CD56^{bright}:CD56^{dim} ratio, decreased cytotoxic activity, and reduced cytokine production (Levy et al., 2011). These abnormalities reflect a suppression of NK cell activity; indeed, many immunotherapeutic approaches to cancer management are targeted at expanding or stimulating the NK cell population (Cheng et al., 2013; Levy et al., 2011). Incidentally, these abnormalities are the opposite of

those seen in the pNK cell population of many women with reproductive failure.

NK cells and infection

Another major function of NK cells is the control of intracellular pathogens including viruses, bacteria, fungi and protozoa. In infection, NK cells are activated either directly or indirectly. They are able to directly recognize and respond to infected cells since viruses tend to downregulate MHC-I molecules and upregulate stress-regulated surface molecules, marking the infected cells as abnormal. Through the indirect pathway, myeloid accessory cells – monocytes, macrophages and DCs – respond to pathogens by releasing cytokines and upregulating costimulatory receptors, which overrides the inhibitory signals emitted by infected but MHC-I-competent cells that would otherwise not be recognized by NK cells. This pathway of indirect activation is especially important in bacterial and protozoal infections and in viruses that have developed strategies to evade direct detection by NK cells (Horowitz, Stegmann, & Riley, 2011). Once activated, NK cells contribute to containing the infection either through cytotoxicity of infected cells or the production of cytokines that control viral replication (Orange & Ballas, 2006). Although pNK cell activity is highest within the first two days of infection, NK cells remain active as the innate immune response transitions to an adaptive one, playing an immunoregulatory role in the later stages of infection (Jost & Altfeld, 2013; Whiteside & Herberman, 1994).

As Orange (2002) notes, one way to appreciate the importance of NK cells in infection is to examine the effects of their absence. Although NK cell deficiencies – numerical or functional – are rare, they are associated with a clear phenotype characterized by increased frequency and severity of infections, especially viral (Whiteside & Herberman, 1994). Susceptibility is increased most significantly to herpes simplex virus (HSV), varicella zoster virus, Epstein–Barr virus (EBV), cytomegalovirus and human papillomavirus, as well as mycobacteria and other bacteria. In genetic disorders resulting in secondary impairment of NK cell cytotoxicity or cytotoxic signalling, patients are most susceptible to infection with HSV and EBV (Orange, 2013). In patients with a complete loss of NK cytotoxicity, often due to perforin deficiency, viral infection can result in overwhelming and potentially fatal disease (Orange & Ballas, 2006). Patients with normal NK cell levels but an overexpression of inhibitory KIRs with strong binding affinity to HLA-C1, such as KIR2DL2, tend to clear acute viruses less quickly and

respond less well to treatment (Horowitz et al., 2011). It follows, therefore, that women with NK cell-related reproductive failure would be less susceptible to viral infections than their immunologically normal or deficient counterparts.

Conclusion

After twenty years of research and debate, it is clear that a significant number of patients with reproductive failure have high NK cell activity. Little is understood, however, of what this finding actually means, and it is not associated with any currently defined disease. Attempts to define a clinical phenotype among these patients should consider the roles of NK cells in other areas of health beyond fertility and the implications of high NK cell activity in these domains. Given what we know about the role of NK cells in non-reproductive disease, it seems likely that NK cell overactivity in the peripheral blood of women with reproductive failure would contribute to health and disease in other respects. Perhaps these women are at higher risk of autoimmune disease, and lower risk of cancer and infection.

Although high NK cell activity appears not to be conducive to reproduction, it also appears to provide protection against other disease. Many reproductive-aged women who are told they have high NK cell activity view their immune system as a curse, when in fact the literature suggests it may be a blessing in other respects. In light of this, it is important to consider the potential ramifications of immunosuppressive therapy in women with high NK cell levels and whether the harm done by such treatments could outweigh the benefits. After years of research and debate focused on reproduction, perhaps it is time to look at the bigger picture of what it means to have elevated NK cell activity.

Disclosure statement

The authors report no conflicts of interest. The authors alone are responsible for the content and writing of the article.

References

- Almasry, S.M., Elmansy, R.A., Elfayomy, A.K., & Algaidi, S.A. (2015). Ultrastructure alteration of decidual natural killer cells in women with unexplained recurrent miscarriage: A possible association with impaired decidual vascular remodelling. *Journal of Molecular Histology*, *46*, 67–78. doi: 10.1007/s10735-014-9598-8.
- Aoki, K., Kajjura, S., Matsumoto, Y., Ogasawara, M., Okada, S., Yagami, Y., & Gleicher, N. (1995). Preconceptional natural-killer-cell activity as a predictor of miscarriage. *Lancet*, *345*, 1340–1342. doi:10.1016/S0140-6736(95)92539-2.
- Beer, A.E., Kwak, J.Y., & Ruiz, J.E. (1996). Immunophenotypic profiles of peripheral blood lymphocytes in women with recurrent pregnancy losses and in infertile women with multiple failed *in vitro* fertilization cycles. *American Journal of Reproductive Immunology*, *35*, 376–382. doi: 10.1111/j.1600-0897.1996.tb00497.x.
- Cheng, M., Chen, Y., Xiao, W., Sun, R., & Tian, Z. (2013). NK cell-based immunotherapy for malignant diseases. *Cellular & Molecular Immunology*, *10*, 230–252. doi: 10.1038/cmi.2013.10.
- Clark, D.A. (2008). Immunological factors in pregnancy wastage: Fact or fiction. *American Journal of Reproductive Immunology*, *59*, 277–300. doi: 10.1111/j.1600-0897.2008.00580.x.
- Clifford, K., Flanagan, A.M., & Regan, L. (1999). Endometrial CD56+ natural killer cells in women with recurrent miscarriage: A histomorphometric study. *Human Reproduction*, *14*, 2727–2730. doi: 10.1093/humrep/14.11.2727.
- Coulam, C.B. & Acacio, B. (2012). Does immunotherapy for treatment of reproductive failure enhance live births? *American Journal of Reproductive Immunology*, *67*, 296–304. doi: 10.1111/j.1600-0897.2012.01111.x.
- Coulam, C.B., Goodman, C., Rousev, R.G., Thomason, E.J., & Beaman, K.D. (1995). Systemic CD56+ cells can predict pregnancy outcome. *American Journal of Reproductive Immunology*, *33*, 40–46. doi: 10.1111/j.1600-0897.1995.tb01136.x.
- Emmer, P.M., Nelen, W.L., Steegers, E.A., Hendriks, J.C., Veerhoek, M., & Joosten, I. (2000). Peripheral natural killer cytotoxicity and CD56(pos)CD16(pos) cells increase during early pregnancy in women with a history of recurrent spontaneous abortion. *Human Reproduction*, *15*, 1163–1169. doi: 10.1093/humrep/15.5.1163.
- Faridi, R.M. & Agrawal, S. (2011). Killer immunoglobulin-like receptors (KIRs) and HLA-C allorecognition patterns implicative of dominant activation of natural killer cells contribute to recurrent miscarriages. *Human Reproduction*, *26*, 491–497. doi: 10.1093/humrep/deq341.
- Flodstrom-Tullberg, M., Bryceson, Y.T., Shi, F.D., Hoglund, P., & Ljunggren, H.G. (2009). Natural killer cells in human autoimmunity. *Current Opinion in Immunology*, *21*, 634–640. doi: 10.1016/j.coi.2009.09.012.
- Fogel, L.A., Yokoyama, W.M., & French, A.R. (2013). Natural killer cells in human autoimmune disorders. *Arthritis Research & Therapy*, *15*, 216. doi: 10.1186/ar4232.
- Folci, M. (2014). Natural killer cells in autoimmunity: Clues or tools? *Inflammation and Cell Signaling*, *1*, e51. doi: 10.14800/ics.51.
- Fukui, A., Funamizu, A., Yokota, M., Yamada, K., Nakamura, R., Fukuhara, R., ... Mizunuma, H. (2011). Uterine and circulating natural killer cells and their roles in women with recurrent pregnancy loss, implantation failure and pre-eclampsia. *Journal of Reproductive Immunology*, *90*, 105–110. doi: 10.1016/j.jri.2011.04.006.
- Fukui, A., Kwak-Kim, J., Ntrivalas, E., Gilman-Sachs, A., Lee, S.K., & Beaman, K. (2008). Intracellular cytokine expression of peripheral blood natural killer cell subsets in women with recurrent spontaneous abortions

- and implantation failures. *Fertility & Sterility*, 89, 157–165. doi: 10.1016/j.fertnstert.2007.02.012.
- Gelbaya, T.A., Potdar, N., Jeve, Y.B., & Nardo, L.G. (2014). Definition and epidemiology of unexplained infertility. *Obstetric & Gynecological Survey*, 69, 109–115. doi: 10.1097/ogx.000000000000043.
- Ghafourian, M., Karami, N., Khodadadi, A., & Nikbakht, R. (2014). Increase of CD69, CD161 and CD94 on NK cells in women with recurrent spontaneous abortion and *in vitro* fertilization failure. *Iranian Journal of Immunology*, 11, 84–96. doi: IJIV11i2A3.
- Hadinedoushan, H., Mirahmadian, M., & Aflatounian, A. (2007). Increased natural killer cell cytotoxicity and IL-2 production in recurrent spontaneous abortion. *American Journal of Reproductive Immunology*, 58, 409–414. doi: 10.1111/j.1600-0897.2007.00524.x.
- Horowitz, A., Stegmann, K.A., & Riley, E.M. (2011). Activation of natural killer cells during microbial infections. *Frontiers in Immunology*, 2, 88. doi: 10.3389/fimmu.2011.00088.
- Imai, K., Matsuyama, S., Miyake, S., Suga, K., & Nakachi, K. (2000). Natural cytotoxic activity of peripheral-blood lymphocytes and cancer incidence: An 11-year follow-up study of a general population. *Lancet*, 356, 1795–1799. doi: 10.1016/s0140-6736(00)03231-1.
- Jost, S. & Altfeld, M. (2013). Control of human viral infections by natural killer cells. *Annual Review of Immunology*, 31, 163–194. doi: 10.1146/annurev-immunol-032712-100001.
- Junovich, G., Azpiroz, A., Incera, E., Ferrer, C., Pasqualini, A., & Gutierrez, G. (2013). Endometrial CD16(+) and CD16(-) NK cell count in fertility and unexplained infertility. *American Journal of Reproductive Immunology*, 70, 182–189. doi: 10.1111/aji.12132.
- Katano, K., Suzuki, S., Ozaki, Y., Suzumori, N., Kitaori, T., & Sugiura-Ogasawara, M. (2013). Peripheral natural killer cell activity as a predictor of recurrent pregnancy loss: A large cohort study. *Fertility & Sterility*, 100, 1629–1634. doi: 10.1016/j.fertnstert.2013.07.1996.
- King, K., Smith, S., Chapman, M., & Sacks, G. (2010). Detailed analysis of peripheral blood natural killer (NK) cells in women with recurrent miscarriage. *Human Reproduction*, 25, 52–58. doi: 10.1093/humrep/dep349.
- Krizhanovsky, V., Yon, M., Dickins, R.A., Hearn, S., Simon, J., Miething, C., ... & Lowe, S.W. (2008). Senescence of activated stellate cells limits liver fibrosis. *Cell*, 134, 657–667. doi: 10.1016/j.cell.2008.06.049.
- Kuroda, K., Venkatakrishnan, R., James, S., Sucurovic, S., Mulac-Jericevic, B., Lucas, E.S., ... & Quenby, S. (2013). Elevated periimplantation uterine natural killer cell density in human endometrium is associated with impaired corticosteroid signaling in decidualizing stromal cells. *Journal of Clinical Endocrinology and Metabolism*, 98, 4429–4437. doi: 10.1210/jc.2013-1977.
- Kwak-Kim, J., Han, A.R., Gilman-Sachs, A., Fishel, S., Leong, M., & Shoham, Z. (2013). Current trends of reproductive immunology practices in *in vitro* fertilization (IVF) - a first world survey using IVF-worldwide.Com. *American Journal of Reproductive Immunology*, 69, 12–20. doi: 10.1111/j.1600-0897.2012.01183.x.
- Laird, S.M., Mariee, N., Wei, L., & Li, T.C. (2011). Measurements of CD56+ cells in peripheral blood and endometrium by flow cytometry and immunohistochemical staining *in situ*. *Human Reproduction*, 26, 1331–1337. doi: 10.1093/humrep/der104.
- Laird, S.M., Tuckerman, E.M., Cork, B.A., Linjawi, S., Blakemore, A.I., & Li, T.C. (2003). A review of immune cells and molecules in women with recurrent miscarriage. *Human Reproduction Update*, 9, 163–174. doi: 10.1093/humupd/dmg013.
- Langers, I., Renoux, V.M., Thiry, M., Delvenne, P., & Jacobs, N. (2012). Natural killer cells: Role in local tumor growth and metastasis. *Biologics*, 6, 73–82. doi: 10.2147/btt.s23976.
- Lee, S.K., Na, B.J., Kim, J.Y., Hur, S.E., Lee, M., Gilman-Sachs, A., & Kwak-Kim, J. (2013). Determination of clinical cellular immune markers in women with recurrent pregnancy loss. *American Journal of Reproductive Immunology*, 70, 398–411. doi: 10.1111/aji.12137.
- Levy, E.M., Roberti, M.P., & Mordoh, J. (2011). Natural killer cells in human cancer: From biological functions to clinical applications. *Journal of Biomedicine and Biotechnology*, 2011, 676198. doi: 10.1155/2011/676198.
- Lu, Y., Zeng, B., Zhang, Y., Xiang, W., Hu, L., & Liao, A. (2011). Quantitative and functional changes in peripheral natural killer cells in women with reproductive failure after artificial insemination with donor sperm. *Journal of Reproductive Immunology*, 91, 83–89. doi: 10.1016/j.jri.2011.05.009.
- Lukassen, H.G., Joosten, I., van Cranenbroek, B., van Lierop, M.J., Bulten, J., Braat, D.D., & van der Meer, A. (2004). Hormonal stimulation for IVF treatment positively affects the CD56bright/CD56dim NK cell ratio of the endometrium during the window of implantation. *Molecular Human Reproduction*, 10, 513–520. doi: 10.1093/molehr/gah067.
- Morikawa, M., Yamada, H., Kato, E.H., Shimada, S., Kishi, T., Yamada, T., ... Fujimoto, S. (2001). Massive intravenous immunoglobulin treatment in women with four or more recurrent spontaneous abortions of unexplained etiology: Down-regulation of NK cell activity and subsets. *American Journal of Reproductive Immunology*, 46, 399–404. doi: 10.1034/j.1600-0897.2001.d01-31.x.
- Ntrivalas, E.I., Kwak-Kim, J.Y., Gilman-Sachs, A., Chung-Bang, H., Ng, S.C., Beaman, K.D., ... Beer, A.E. (2001). Status of peripheral blood natural killer cells in women with recurrent spontaneous abortions and infertility of unknown aetiology. *Human Reproduction*, 16, 855–861. doi: 10.1093/humrep/16.5.855.
- Orange, J.S. (2002). Human natural killer cell deficiencies and susceptibility to infection. *Microbes and Infection*, 4, 1545–1558. doi: 10.1016/S1286-4579(02)00038-2.
- Orange, J.S. (2013). Natural killer cell deficiency. *Journal of Allergy and Clinical Immunology*, 132, 515–526. doi: 10.1016/j.jaci.2013.07.020.
- Orange, J.S. & Ballas, Z.K. (2006). Natural killer cells in human health and disease. *Clinical Immunology*, 118, 1–10. doi: 10.1016/j.clim.2005.10.011.
- Ozcimen, E.E., Kiyici, H., Uckuyu, A., & Yanik, F.F. (2009). Are CD57+ natural killer cells really important in early pregnancy failure? *Archives of Gynecology and Obstetrics*, 279, 493–497. doi: 10.1007/s00404-008-0736-y.
- Polanski, L.T., Barbosa, M.A., Martins, W.P., Baumgarten, M.N., Campbell, B., Brosens, J., ... Raine-Fenning, N. (2014). Interventions to improve reproductive outcomes in women with elevated natural killer cells undergoing

- assisted reproduction techniques: A systematic review of literature. *Human Reproduction*, 29, 65–75. doi: 10.1093/humrep/det414.
- Quenby, S., Kalumbi, C., Bates, M., Farquharson, R., & Vince, G. (2005). Prednisolone reduces preconceptual endometrial natural killer cells in women with recurrent miscarriage. *Fertility & Sterility*, 84, 980–984. doi: 10.1016/j.fertnstert.2005.05.012.
- Quenby, S. & Farquharson, R. (2006). Uterine natural killer cells, implantation failure and recurrent miscarriage. *Reproductive BioMedicine Online*, 13, 24–28. doi: 10.1016/S1472-6483(10)62012-3.
- Quenby, S., Nik, H., Innes, B., Lash, G., Turner, M., Drury, J., & Bulmer, J. (2009). Uterine natural killer cells and angiogenesis in recurrent reproductive failure. *Human Reproduction*, 24, 45–54. doi: 10.1093/humrep/den348.
- Ramos-Medina, R., Garcia-Segovia, A., Leon, J.A., Alonso, B., Tejera-Alhambra, M., Gil, J., ... Sanchez-Ramon, S. (2013). New decision-tree model for defining the risk of reproductive failure. *American Journal of Reproductive Immunology*, 70, 59–68. doi: 10.1111/aji.12098.
- Roussev, R.G., Kaider, B.D., Price, D.E., & Coulam, C.B. (1996). Laboratory evaluation of women experiencing reproductive failure. *American Journal of Reproductive Immunology*, 35, 415–420. doi: 10.1111/j.1600-0897.1996.tb00503.x.
- Sacks, G. (2014). *NK cells in peripheral blood and the endometrium*. In Christiansen, O.B. (ed), *Recurrent Pregnancy Loss*. West Sussex, UK: John Wiley & Sons.
- Sacks, G. (2015). Enough! stop the arguments and get on with the science of natural killer cell testing. *Human Reproduction*, 30, 1526–1531. doi: 10.1093/humrep/dev096.
- Sacks, G., Yang, Y., Gowen, E., Smith, S., Fay, L., & Chapman, M. (2012). Detailed analysis of peripheral blood natural killer cells in women with repeated IVF failure. *American Journal of Reproductive Immunology*, 67, 434–442. doi: 10.1111/j.1600-0897.2012.01105.x.
- Santillan, I., Lozano, I., Illan, J., Verdu, V., Coca, S., Bajo-Arenas, J.M., & Martinez, F. (2015). Where and when should natural killer cells be tested in women with repeated implantation failure? *Journal of Reproductive Immunology*, 108, 142–148. doi: 10.1016/j.jri.2014.12.009.
- Simon, A. & Laufer, N. (2012). Assessment and treatment of repeated implantation failure (RIF). *Journal of Assisted Reproduction and Genetics*, 29, 1227–1239. doi: 10.1007/s10815-012-9861-4.
- Terunuma, H., Deng, X., Dewan, Z., Fujimoto, S., & Yamamoto, N. (2008). Potential role of NK cells in the induction of immune responses: Implications for NK cell-based immunotherapy for cancers and viral infections. *International Reviews of Immunology*, 27, 93–110. doi: 10.1080/08830180801911743.
- Thum, M.Y., Bhaskaran, S., Bansal, A.S., Shehata, H., Ford, B., Sumar, N., & Abdalla, H.I. (2005). Simple enumerations of peripheral blood natural killer (CD56+ NK) cells, B cells and T cells have no predictive value in IVF treatment outcome. *Human Reproduction*, 20, 1272–1276. doi: 10.1093/humrep/deh774.
- Tuckerman, E., Laird, S.M., Prakash, A., & Li, T.C. (2007). Prognostic value of the measurement of uterine natural killer cells in the endometrium of women with recurrent miscarriage. *Human Reproduction*, 22, 2208–2213. doi: 10.1093/humrep/dem141.
- Varla-Leftherioti, M., Spyropoulou-Vlachou, M., Keramitsoglou, T., Papadimitropoulos, M., Tsekoura, C., Graphou, O., ... & Stavropoulos-Giokas, C. (2005). Lack of the appropriate natural killer cell inhibitory receptors in women with spontaneous abortion. *Human Immunology*, 66, 65–71. doi: 10.1016/j.humimm.2004.10.005.
- Whiteside, T.L. & Herberman, R.B. (1994). Role of human natural killer cells in health and disease. *Clinical and Diagnostic Laboratory Immunology*, 1, 125–133. <http://www.ncbi.nlm.nih.gov/pmc/articles/PMC368214/>.
- Yamamoto, T., Takahashi, Y., Kase, N., & Mori, H. (1999). Decidual natural killer cells in recurrent spontaneous abortion with normal chromosomal content. *American Journal of Reproductive Immunology*, 41, 337–342. doi: 10.1111/j.1600-0897.1999.tb00447.x.
- Yoo, J.H., Kwak-Kim, J., Han, A.R., Ahn, H., Cha, S.H., Koong, M.K., ... Yang, K.M. (2012). Peripheral blood NK cell cytotoxicities are negatively correlated with CD8(+) T cells in fertile women but not in women with a history of recurrent pregnancy loss. *American Journal of Reproductive Immunology*, 68, 38–46. doi: 10.1111/j.1600-0897.2012.01133.x.